

## ***Errata to the lab manual***

Chemistry 184, Biological Chemistry Laboratory, Spring 2008

### DETECTION OF FLUORESCENCE FROM SINGLE MOLECULES

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Page 38 – **Fine alignment of the excitation laser.**

*Current protocol:* Perform steps 1-13 using the 60x objective.

*Corrected protocol:* First perform steps 1-13 using the 10x objective. Then, repeats steps 1-13 using the 60x objective.

### CHARACTERIZATION OF MUSHROOM TYROSINASE ACTIVITY

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Page 83 – **Step 2: Packing the column with flow.** The protocol is missing step #14.

14. Detach the column without introducing air onto the column. Perform step #1 from the *Protocol for attaching a column or introducing a new buffer*, page 85.

Page 87 – **Step 4: Eluting your sample from the column**, step #6.

*Current protocol:* “Three CV is a good volume for this gradient...”

*Corrected protocol:* “Five CV is a good volume for this gradient...”

Page 92 – **Step 1: Concentrating the samples**, step #6.

*Current protocol:* “Add enough *DEAE loading buffer* to bring all of your samples up to 50  $\mu\text{l}$ .”

*Corrected protocol:* “If necessary, add enough *DEAE loading buffer* to bring all of your samples up to 25  $\mu\text{l}$ .”

### ALTERATION OF THE SPECTRAL PROPERTIES OF ZSYELLOW

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Page 121 – **Purification of His-tagged protein**, steps #5 and #6.

Steps #5 and #6 should be reversed.

## APPENDICES

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### Page 141 – Step 1: Making the dilution series.

*Current protocol:* the last column reads

BSA ( $\mu\text{L}$ ) (stock or dilution)
100 of BSA stock
100 of dilution 2
100 of dilution 3
100 of dilution 4
100 of dilution 5
100 of dilution 6
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*Corrected protocol:* the last column reads

BSA ( $\mu\text{L}$ ) (stock or dilution)
100 of BSA stock
100 of dilution 1
100 of dilution 2
100 of dilution 3
100 of dilution 4
100 of dilution 5
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## RECIPES

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### Page 147 – Buffers and Salts for Tyrosinase Experiment, DEAE elution buffer.

The buffer should contain 1 M sodium chloride (*not* 25 mM sodium chloride). When making 500 mL of buffer, 29.2 g of sodium chloride is included.

### Page 148 – Buffers for Protein Gels, 10x SDS-PAGE running buffer

The buffer should contain 740 mM glycine (*not* 2 M glycine). 72.1 g is the correct amount of glycine to add.